Genomic DNA

User manuals

NucleoMag® DNA Microbiome 744330.1 / .4

Dear valued customer,

Thank you for choosing MACHEREY-NAGEL for your application. We have attached a short protocol for your review.

To obtain the best results we recommend to follow the detailed protocol available online, especially when you are a first time user of this kit.

All important links to the above mentioned product are listed in this leaflet.

Important information regarding product components, specifications, safety instructions, and processing protocols, can all be found on the product website and accessed easily via the QR code.

QR-Code product website

qr.mn-net.com/qr/(241)744330

Use the following QR code or the link below for direct access to the user manual.

QR-Code user manual

qr.mn-net.com/qr/(IFU)744330

It is strongly recommended to read the detailed protocol section of the user manual if using the kit for the first time. However, experienced users may refer to the protocol at a glance. The protocol at a glance is designed to be used only as a supplemental tool for quick reference while performing the purification procedure.

We are constantly improving our products and we reserve the right to make changes or additions to protocols. Please check for updated revisions for previously downloaded manuals.

This leaflet does not replace the full manual!

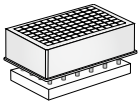
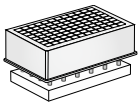
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Protocol at a glance

Isolation of DNA from soil, stool and biofilm samples using MN Bead Tubes

1	**Prepare sample**	MN Bead Tube Type A Up to 200 mg sample material 700 µL MI1 *Optional (Soil): 150 µL Enhancer SX* *Optional: 2.5 µL Liquid RNase A* *Optional (Stool / Biofilm): 70 °C, 5 min* Agitate on a swing mill or similar device for 1 – 10 min
2	**Precipitate debris**	Incubate for 5 min at RT 11,000 x *g*, 5 min Transfer supernatant to a fresh tube
3	**Precipitate contaminants**	150 µL MIc Vortex 5 s 10 min, 2 – 8 °C 11,000 x *g*, 5 min Transfer supernatant into a fresh tube
4	**Bind DNA to NucleoMag® B-Beads**	Up to 500 µL lysate 25 µL NucleoMag® B-Beads 310 µL MI2
		Mix by shaking for 5 min at RT (Optional: Mix by pipetting up and down)
		Remove supernatant after 5 min separation
5	**Wash with MI3**	Remove Square-well Block from NucleoMag® SEP 600 µL MI3
		Resuspend: Shake 2 min at RT (Optional: Mix by pipetting up and down)
		Remove supernatant after 2 min separation
6	**Wash with MI3**	Remove Square-well Block from NucleoMag® SEP 600 µL MI3
		Resuspend: Shake 2 min at RT (Optional: Mix by pipetting up and down)
		Remove supernatant after 2 min separation
7	**Wash with MI4**	Remove Square-well Block on NucleoMag® SEP 600 µL MI4
		Resuspend: Shake 2 min at RT (Optional: Mix by pipetting up and down)
		Remove supernatant after 2 min separation
8	**Wash with 70 % EtOH**	Remove Square-well Block on NucleoMag® SEP 600 µL 70 % EtOH
		Resuspend: Shake 2 min at RT (Optional: Mix by pipetting up and down)
		Remove supernatant after 2 min separation

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9 Dry the beads		10 – 15 min at RT
10 Elute DNA		Remove Square-well Block from NucleoMag® SEP 50 – 100 µL MI5
		Shake 5 min at RT (Optional: Mix by pipetting up and down)
		Separate 2 min and transfer DNA into elution plate

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QR-Code Safety Data Sheet

Please find a digital version of the safety data sheets by following the link below:

www.mn-net.com/sds

We strongly recommend to carefully read the detailed protocol section of the product's user manual. If you have any questions about the protocol or product, please contact our Technical Support.

Contact MN

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