

## Specification

Solid medium for the enumeration of heterotrophic microorganisms in treated waters according to Pharmacopoeial Method.

## Presentation

30 Prepared plates

55 mm Plates for filtration purposes  
with:  $9 \pm 1$  ml

### Packaging Details

1 box containing: 5 plastic bags with 6 plates of 55 mm/ bag.

### Shelf Life

6 months

### Storage

2-25°C

## Composition

Composition (g/l):

Proteose peptone..... 0.500

Casein hydrolysate (Tryptone)..... 0.500

Yeast extract..... 0.500

D(+)-Glucose..... 0.500

Starch..... 0.500

Sodium pyruvate..... 0.300

Dipotassium hydrogen phosphate. 0.300

Magnesium sulphate (anhydrous). 0.024

Agar..... 15.000

## Description /Technique

### Description

R2A Agar was proposed in 1979 by Reasoner and Geldenreich and a few years later accepted by the APHA as an alternative medium for the enumeration of stressed cells in treated potable water. The culture medium has also been adopted by the European Pharmacopoeia for the control of purified water.

The use of nutrient rich media like PCA or TSA allows the growth of most microbes, but does not permit the recuperation of stressed or chlorine resistant organisms. Using a medium like R2A with low nutrients in combination with a lower temperature and longer incubation time it is possible to induce the resuscitation of these damaged cells.

In R2A Agar the source of nitrogen is the peptone and Yeast Extract supplies the vitamins and growth factors. The source of carbon is dextrose and magnesium sulfate and potassium phosphate maintain the osmotic pressure. The starch is a detoxifier and sodium pyruvate increases the recuperation of stressed cells. The agar acts as gelling agent.

### Technique

The water sample must be processed as quickly as possible. If it is not possible to process within the first 6 hours, the sample must be refrigerated, but not for more than 30 hours.

Collect, dilute and prepare samples and volumes to be filtered as required according to specifications, directives, official standard regulations and/or expected results.

Filter the sample through a 0.45 mm pore membrane and apply it onto the surface of the agar.

When incubating at 35°C, an incubation period of 3-5 days is recommended. In most circumstances an incubation temperature of 20-25°C for 5-7 days is more effective. Plates must be protected against dehydration.

After incubation, enumerate all the colonies that have appeared onto the surface of the membrane.

Calculate total microbial count per ml of sample by multiplying the average number of colonies per plate by the inverse dilution factor. Report results as Colony Forming Unit (CFU's) per ml along with incubation time and temperature.

**Quality control****Physical/Chemical control**

Color : Pale yellow

pH: 7.2 ± 0.2 at 25°C

**Microbiological control**

Growth Promotion Test according to harmonized pharmacopoeial monographs and test methods &amp; ISO 11133:2014

Membrane Filtration /Practical range 100±20 CFU; Min. 50 CFU (Productivity)./10<sup>4</sup>-10<sup>6</sup> CFU for Selectivity.

Aerobic.Incubation at 30-35°C for 24-48h (bacteria) and 20-25°C for 3-5 days (moulds and yeast).

**Microorganism****Growth***Ps. aeruginosa* ATCC® 9027, WDCM 00026

Good (≥70 %)

*Bacillus subtilis* ATCC® 6633, WDCM 00003

Good (≥70 %)

*Escherichia coli* ATCC® 8739, WDCM 00012

Good (≥70 %)

*Aspergillus brasiliensis* ATCC® 16404, WDCM 00053

Good (≥70 %)

*Candida albicans* ATCC® 10231, WDCM 00054

Good (≥70 %)

*Staphylococcus aureus* ATCC® 6538, WDCM 00032

Good (≥70 %)

**Sterility Control**

Incubation 48 hours at 30-35°C and 48 hours at 20-25°C: NO GROWTH

Check at 7 days after incubation in same conditions

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