

**Specification**

Selective and differential medium for the detection and enumeration of coliforms and *E. coli* in water samples by MF technique.

**Presentation**

20 Prepared Plates  
90 mm  
with: 21 ± 2 ml

**Packaging Details**

1 box with 2 packs of 10 plates/pack. Single cellophane.

**Shelf Life**

3 months

**Storage**

2-14°C

**Composition**

Composition (g/l):

Enzymatic digest of casein.....	1.00
Yeast extract.....	2.00
Sodium chloride.....	5.00
Di-sodium hydrogen phosphate.....	2.70
Sodium dihydrogen phosphate dihydrate.....	2.20
Tryptophan.....	1.00
Sodium pyruvate.....	1.00
Tergitol®7.....	0.15
Sorbitol.....	1.00
6-Chloro-3-indoxyl- β-D-galactopyranoside.....	0.20
5-Bromo-4-chloro-3- indoxyl-β-D-glucuronic acid.....	0.10
IPTG.....	0.10
Agar.....	13.00

## Description /Technique

### Description:

The combined action of peptone, pyruvate and sorbitol allow rapid colony growth in this phosphate buffered medium, which also permits simple recovery of sublethal thermally injured coliforms. Sodium chloride provides the correct osmotic environment necessary for growth.

The selectivity is attained, partially, by the Tergitol<sup>®</sup> 7, which inhibits the growth of Gram positive bacteria and some Gram negative without effecting the coliform bacteria. The colonial differentiation is due to the chromogenic mixture, composed of two enzyme substrates: 6-chloro-3-indoxyl-β-D-galacto-pyranoside (Salmon<sup>®</sup>-GAL) and 5-bromo-4-chloro-3-indoxyl-β-D-glucuronide (X-Glucuronide). The first one is cleaved by the characteristic enzyme found in coliforms, β-D-galactosidase and gives a salmon-red colour to the coliform colonies. The second chromogenic substance is cleaved by the β-D-glucuronidase enzyme characteristic of *E. coli* and turns the colonies of these bacteria a blue colour. *E. coli* has the two enzymes and cleaves both chromogenic substances giving dark blue to violet colonies. Total coliforms are the sum of *E. coli* colonies plus salmon-red colonies. The IPTG, enhances the reactions described above. Other Gram negative bacteria produce colourless colonies except some that possess glucuronidase activity (but not galactosidase) and they produce light blue to turquoise colonies.

To confirm the *E. coli* colonies in this medium a small amount of tryptophane is included verifying indol production: coat the blue-violet colonies with a drop of Kovacs Reagent. If the reagent turns a cherry-red colour in a few seconds this confirms the production of indol and hence the presence of *E. coli*. When the Chromogenic Agar for Coliform is used with the membrane filter method, the colour and growth of the colonies can be modified by the characteristics of the membrane filter. It is advisable to perform validation of the membrane filter type used. The Spanish Health Ministry (Ministerio de Sanidad y Consumo) has officially adopted this medium as an alternative methodology for the microbiological analysis of water for human consumption, giving a new definition for *Escherichia coli* ("Enterobacteriaceae that express the β-D- galactosidase and the β-D-glucuronidase enzymes simultaneously") and coliform bacteria: "Enterobacteriaceae that express the β-D-galactosidase enzyme".

### Limitation of the procedure:

The production of β-galactosidase, although common to all the coliforms, varies from one strain to another being influenced by the temperature and incubation time. At temperatures above 37 ° C its production decreases, causing a loss of reddish color intensity, while the bluish tones in the strains of *Escherichia coli* are accentuated.

If the membrane filtration method is used, it must be taken into account that the nature and characteristics of the filter membrane used also influences the size and color of the colonies grown on this culture medium.

### Technique:

The water sample is filtered through a membrane filter of 0,45 μm of pore diameter validated according to the ISO Standard 7704:1985. The membrane is then placed on the surface of the CCA medium avoiding entrapment of air bubbles between the membrane and agar surface. The petri dish with the membrane is incubated for 18-24 hours at 36 ± 2°C. If in 18 h there is growth of red or colourless colonies, extend the incubation until 24 h to include late reactions of β-galactosidase or β-glucuronidase. Count β-galactosidase positive colonies and β-glucuronidase negative colonies (all colonies coloured from salmon-rose to red) as Coliform bacteria not *E. coli*. Count β-galactosidase positive colonies and β-glucuronidase positive colonies (all colonies coloured from deep blue to violet) as *E. coli*. Total Coliform count is obtained by the addition of the salmon-rose to red colonies plus the deep blue to violet colonies.

## Quality control

### Physical/Chemical control

Color : Pale yellow                      pH: 6.8 ± 0.2 at 25°C

### Microbiological control

Membrane Filtration /Practical range 100±20 CFU; Min. 50 CFU (Productivity)./10<sup>4</sup>-10<sup>6</sup> CFU for Selectivity. Microbiological control according to ISO 11133:2014/ Adm 1:2018.

Aerobiosis. Incubation at 36 ± 2°C, reading at 18-24 h

### Microorganism

*Escherichia coli* ATCC<sup>®</sup> 25922, WDCM 00013

*Escherichia coli* ATCC<sup>®</sup> 8739, WDCM 00012

*Citrobacter freundii* ATCC<sup>®</sup> 43864, WDCM 00006

*Ps. aeruginosa* ATCC<sup>®</sup> 10145, WDCM 00024

*Enterococcus faecalis* ATCC<sup>®</sup> 19433, WDCM 00009

### Growth

Good (≥70%) Dark-blue to violet colonies

Good (≥70%) Dark-blue to violet colonies

Good (≥70%) Salmon to red colonies

Good - Colourless colonies

Inhibited

### Sterility Control

Incubation 48 hours at 30-35°C and 48 hours at 20-25°C: NO GROWTH

Check at 7 days after incubation in same conditions

**Bibliography**

- ADAMS, M., R.GRUBB, S.M. HAMER & A. CLIFFORD (1990) Colorimetric enumeration of *Escherichia coli* based on  $\beta$ -glucuronidase activity. *Appl. Environ. Microbiol.* 56:2021.
- ISO 7704 Standard (1985) Water Quality - Evaluation of membrane filters used for microbiological analyses.
- ISO 9308-1: 2014/Amd.1:2016(E) Water quality. Enumeration of *Escherichia coli* and coliform bacteria - Part 1: Membrane filtration method for waters with low bacterial background flora.
- ISO 11133:2014/ Adm 1:2018. Microbiology of food, animal feed and water. Preparation, production, storage and performance testing of culture media.
- KILIAN, M. & P. BÜLOW (1976) Rapid Diagnostic of Enterobacteriaceae. I. Detection of bacterial glycosidases. *Acta Pathol. Microbiol. Scand. Sect. B* 84:245-251.
- MANAFI, M & W. KNEIFEL (1989) A combined chromogenic-fluorogenic medium for the simultaneous detection of total coliform and *E. coli* in water. *Zentralbl. Hyg.* 189:225-234.
- UNE-EN ISO 11133 (2014). Microbiología de los alimentos para consumo humano, alimentación animal y agua.-Preparación, producción, conservación y ensayos de rendimiento de los medios de cultivo.
- TURNER, K.M., L. RESTAINO & E.W. FRAMPTON (2000) Efficacy of Chromocult Coliform Agar for coliform and *Escherichia coli* detection in Foods. *J.Food Protect.* 63(4):539-541