

**Specification**

Liquid medium for the selective enrichment of *Salmonella* in foodstuffs and other samples, according to ISO and FIL-IDF standards.

**Presentation**

20 Tubes  
Tube 16 x 113 mm  
with: 10 ± 0,2 ml

**Packaging Details**

16x113 mm glass tubes, ink labelled, metal-Non  
injectable cap. - 20 tubes per box

**Shelf Life**

10 months

**Storage**

8-25°C

**Composition**

Composition (g/l):

Soy peptone..... 4.500  
Sodium chloride..... 7.200  
Monopotassium phosphate..... 1.260  
Dipotassium phosphate..... 0.180  
Magnesium chloride anhydrous.... 13.400  
Malachite Green..... 0.036

**Description /Technique**Description:

The Rappaport Vassiliadis medium complies with the recommendations of the APHA for the examination of food.

This culture medium is a modification of the R10 Medium (from Rappaport *et al.*) or RV Broth (from Vassiliadis *et al.*) by van Schothorst & Renaud. The modifications are an adjustment in the magnesium chloride concentration and the buffering capacity of the medium to aid pH maintenance during storage. It shows a higher selectivity towards *Salmonella* and produces better yields than other similar media, especially after preliminary enrichment and at an incubation temperature of 41 ± 0,5°C.

Malachite green, low pH and magnesium chloride inhibit the growth of microorganisms normally found in the intestine but do not affect the proliferation of most salmonellae. As malachite green inhibits the growth of *Shigella*, other culture methods may need to be used to isolate this organism. The addition of soy peptone enhances the growth of *Salmonella*.

Technique:

Collect, dilute and prepare samples and volumes as required according to specifications, directives, official standard regulations and/or expected results.

Inoculate aseptically the tubes with the prepared sample or its dilution in a proportion 1/10 (V/V).

Incubate the tubes tightly closed aerobically at 41 ± 0,5 ° C for 24h ± 3 h..

(Incubation times, temperature and sample volumes may vary depending on the sample, on the specifications,...)

Read the turbidity (growth indicator) and inoculate any confirmatory, secondary medium by streaking methodology or by spiral method, like XLD, BPLS,... to confirm results after proper incubation, enumerate all the colonies that have appeared onto the surface of the secondary agar.

Presumptive isolation / recovery of *Salmonella* must be confirmed by further microbiological and biochemical tests.

Each laboratory must evaluate the results according to their specifications.

Calculate total microbial count per ml of sample by multiplying the average number of colonies per plate by the inverse dilution factor if streaked a diluted sample. Report results as Colony Forming Unit (CFU's) per ml or g along with enrichment and secondary media used, incubation time and temperature.

**Quality control****Physical/Chemical control**

Color : Blue

pH: 5.2 ± 0.2 at 25°C

**Microbiological control**Inoculate: Practical range 100 ± 20 CFU; Min. 50 CFU (Productivity)/ 10<sup>4</sup>-10<sup>6</sup> (Selectivity).

Microbiological control according to ISO 11133:2014/ Adm 1:2018.

Aerobiosis. Incubation 41,5±1 °C, reading 24h ± 3h. Confirm in XLD (Pr) y TSA (Select.)

**Microorganism***Enterococcus faecalis* ATCC® 29212, WDCM 00087*Escherichia coli* ATCC® 25922, WDCM 00013*S. typhimurium* (14028) + *E. coli* (8739) + *Ps.* (27853)**Growth**

Inhibited. Confirm in TSA at 37°C±1 reading 24 ± 3h

Partial Inhibition. Confirm in TSA at 37°C±1 reading 24 ±

*Salmonella* coln. charact. in XLD (37°C±1 / 24 ± 3h) ≥ 10**Sterility Control**

Incubation 48 hours at 30-35°C and 48 hours at 20-25°C: NO GROWTH

Check at 7 days after incubation in same conditions

**Bibliography**

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