

## Specification

Solid differential medium for primary identification of enterobacteria based on the fermentation of two sugars and the hydrogen sulfide production according to ISO standard.

## Presentation

20 Tubes / Slant  
Tube 16 x 110mm  
with: 7,5 ± 0,3 ml

### Packaging Details

1 box with 20 tubes, 16x113 mm glass tubes, ink labelled and metal cap.

### Shelf Life

9 months

### Storage

8-25°C

## Composition

Composition (g/l):

Peptone.....	20.0
Meat extract.....	3.00
Yeast extract.....	3.00
Sodium chloride.....	5.00
Lactose.....	10.0
Glucose.....	1.00
Ferric citrate.....	0.50
Sodium thiosulfate.....	0.50
Phenol red.....	0.03
Agar.....	15.0

## Description /Technique

Kligler Agar is a differential medium that has all the characteristics of the 2-Sugar Russell Agar and Lead Acetate Medium for H<sub>2</sub>S detection. In this medium, lactose fermentation and hydrogen sulfide production can be detected, allowing a presumptive identification of most enterobacteria. Sugar fermentation is shown by acid production, which turns the indicator from red to yellow. Since there is only a small amount of sugar (dextrose) in the medium, acid production due to its fermentation is very limited and re-oxidation of the indicator occurs on the surface of the medium, causing the indicator to remain red. When lactose is fermented, a large amount of acid is produced re-oxidation does not occur and the entire medium turns yellow.

Hydrogen sulfide production is indicated by the medium turning black, due to the reaction of H<sub>2</sub>S (liberated from thiosulfate) with the iron ions presents in the ammonium iron citrate.

Kligler Iron Agar is used in slanted tubes with short slant and a generous butt, which are inoculated on the surface and also stab inoculated. The inoculum must be copious; it has to come from a solid medium, otherwise, readings may be delayed (up to additional 2-3 days). Normal incubation is 18-24 hours at 36°C ±0,2.

A large production of H<sub>2</sub>S may make the readings difficult, and hence early readings are strongly recommended.

To inoculate tubes follow the standard laboratory methods or the applicable norms:stab inoculation, loop inoculation.

## Quality control

### Physical/Chemical control

Color : Reddish pH: 7.4 ± 0.2 at 25°C

### Microbiological control

Inoculate by stabbing the butt + streak the slant

Aerobiosis. Incubation at 36 ± 2°C, reading at 18-24 h

### Microorganism

*Shigella flexneri* ATCC® 12022, WDCM 00126

*Escherichia coli* ATCC® 8739, WDCM 00012

*Salmonella typhimurium* ATCC® 14028, WDCM 00031

*Proteus mirabilis* ATCC® 43071

### Growth

Good /Slant: Alk/Butt:Ac /Gas (-)/ SH2(-)

Good / Slant:Ac /Butt:Ac /Gas (+)/ SH2(-)

Good /Slant: Alk/Butt:Ac /Gas (+)/ SH2(+)

Good /Slant: Alk/Butt:Ac /Gas (+)/ SH2(+)

### Sterility Control

Incubation 48 hours at 30-35°C and 48 hours at 20-25°C: NO GROWTH

Check at 7 days after incubation in same conditions

**Bibliography**

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