

Specification

Selective supplement for the isolation of pathogenic Enterobacteria, especially *Salmonella*.

Presentation

1 Prepared bottle
Bottle 125 ml
with: 100 ± 3 ml

Packaging Details

1 box with 1 bottle 125 ml. Injectable cap: Plastic screw inner cap. The use of syringes needles with a diameter greater than 0.8 mm is not recommended.

Shelf Life

16 months

Storage

8-25 °C

Composition

Composition (%/vial):

Tergitol 4..... 26

Description /Technique

Description

XLT4 Agar Base (Cat. 1159) with Tergitol 4 supplement, was developed in 1990 by Miller and Tate. It is a highly selective medium for isolating *Salmonella* from competing bacteria such as *Proteus*. Tergitol 4 inhibits the growth of those non-*Salmonella* organisms.

They reported the isolation of non-typhi *Salmonella* from chicken and farm environmental drag-swab samples from heavily contaminated samples. XLT4 Agar can be used clinically to screen stool samples for non-typhoid *Salmonella*.

This medium allows the optimum growth of *Salmonella*. Differentiation of *Salmonella* from other organisms in this medium is based on the fermentation of carbohydrates (Lactose, Xylose, Sucrose) with the resulting production of hydrogen sulfide. H₂S production is detected by the reaction of the iron salt, colonies appearing black or black-centered. Sodium thiosulfate and ferric ammonium citrate are the H₂S indicators. The bacteria that decarboxylate the L-Lysine to cadaverine are identified by the presence of a purple-red color around the colonies due to the elevation of the pH. Phenol red is the pH indicator. Sodium thiosulfate is also added as a source of inorganic sulfur. Yeast extract and peptone are a nitrogen and amino acids source. Bacteriological agar is the solidifying agent.

Typical *Salmonella* colonies (H₂S-positive) appear black or black-centered with a yellow halo after 18-48 hours of incubation at a temperature of 35±2 °C. Upon continued incubation, the colonies become entirely black or pink to red with black centers. Colonies of H₂S-negative *Salmonella* strains appear pink-yellow.

Most *Citrobacter* colonies are yellow without evidence of blackening. The growth of *Enterobacter aerogenes* and *Escherichia coli* is markedly inhibited; colonies that grow in this medium appear yellow without evidence of blackening. The growth of *Proteus*, *Pseudomonas* and *Yersinia enterocolitica* is markedly to completely inhibited. *Shigella* species are partially inhibited and colonies appear red.

Technique:

Add 4,6 ml of XLT4 supplement to 1 L of XLT4 Agar Base (Cat. 1159). Mix well and heat with frequent agitation until completely dissolved. Boil for one minute. AVOID OVERHEATING. DO NOT AUTOCLAVE. Distribute into sterile containers.

Instructions for use:

- Inoculate the sample in a pre-enrichment medium, such as Tetrathionate Broth (Cat. 1114).
- Incubate at 35±2 °C for 18-24 hours.
- Spread or streak the sample from the enrichment medium on the surface of the XLT4 Agar Base.
- Incubate aerobically at a temperature of 35±2 °C for 18-48 hours.

Quality control**Physical/Chemical control**

Color : Colorless - light yellow / transparent pH: at 25°C

Microbiological control

Distribute the complete medium, cooled at 50°C, in plates

Inoculate: Practical range 100 ± 20 CFU. min. 50 CFU (productivity)/ 10⁴-10⁶ (selectivity).

Analytical methodology according to ISO 11133:2014/A1:2018; A2:2020.

Aerobiosis. Incubation at 35 ± 2 °C, reading at 18-48 h

Microorganism

Salmonella enterica ATCC® 13076, WDCM 00030

Salmonella typhimurium ATCC® 14028, WDCM 00031

Escherichia coli ATCC® 25922, WDCM 00013

Enterobacter aerogenes ATCC® 13048, WDCM 00175

Proteus mirabilis ATCC® 29906

Shigella flexneri ATCC® 12022, WDCM 00126

Enterococcus faecalis ATCC® 29212, WDCM 00087

Sterility Control

Incubation 48 h at 30-35 °C and 48 h at 20-25 °C: NO GROWTH.

Check at 7 days after incubation in same conditions.

Growth

Good - Red colonies, black center and SH2 (+)

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Moderate growth - Yellow colonies

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Poor

Dark pink to reddish colonies

Inhibited

Bibliography

Miller, R. G., and C. R. Tate. 1990. XLT4: A highly selective plating medium for the isolation of Salmonella. The Maryland Poultryman, April:2-7.

Tate, C. R., R. G. Miller, and E. T. Mallinson. 1992. Evaluation of two isolation and two non-isolation methods for detecting naturally occurring salmonellae from broiler flock environmental drag-swab samples. J. Food Prot. 55:964-967.

Dusch, H., and M. Altwegg. 1995. Evaluation of five new plating media for the isolation of Salmonella species. J. Clin. Microbiol. 33:802-804.