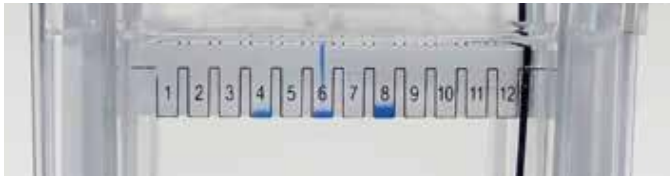
5. Please use a standard 10 µl pipette to load your sample. Please load the sample by inserting the pipette tip vertically into the well.
Maximal volume per well is 60 µl.



6. Electrophoresis Condition

Voltage	Starting current	Finished current	Run Time per Gel*
140 V	75 - 100 mA	30 - 50 mA	45 - 55 min

**Running time is dependent on expected protein sizes, gel percentage and power supply used.*

7. Compatibility

8 × 10 cm	10 × 10 cm
- FastGene® Protein Chamber (NG-002) - Bio-Rad Mini-PROTEAN® II & 3 & Tetra System; - Hoefer SE250 Mighty Small II Mini & SE260 Mighty Small II Deluxe	- LONZA PAGEr™ Minigel Chamber, - Hoefer SE260 Mighty Small II Deluxe, - Life Technologies Novex XCell Surelock® & Bolt™ Mini Gel Tank*

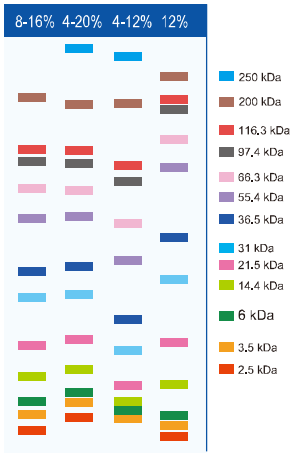
**Extra cushion needed*

8. Opening a gel cassette with an FastGene® Opener

QuickGuide for FastGene® PAGE Gels 8 × 10 cm

NIPPON Genetics EUROPE
Binsfelder Straße 77 | D-52351 Dueren | Germany

1. Choose the appropriate gels for your protein electrophoresis analysis using the charts given below.



Percentage	Seperation range
8-16 %	10 kDa to 200 kDa
4-20 %	10 kDa to 250 kDa
4-12 %	20 kDa to 250 kDa
12 %	6.5 kDa to 200 kDa

2. Preparation of sample:

Reagent	Volume
Protein Sample*	x µl
Deionized H_2O	Up to 8 µl
5 x Loading buffer	2 µl
Total volume**	10 µl

*Heat the sample at 100 °C for 10 min (not for native gels).
**Maximal volume per well is 60 µl.

3. Prepare the gel tank and the running buffer. Please use FastGene® MOPS (PG-MOPS10) or MES buffer as a PAGE running buffer. Do not use Tris-Glycine.

NOTE: The FastGene® MOPS (PG-MOPS10) buffer contains SDS and is therefore not suitable for native PAGE.

MOPS Buffer	
Tris-base	6.06 g
MOPS	10.46 g
EDTA	0.3 g
(SDS)*	(1 g)*
H_2O	up to 1000 ml

*Not for native gels

MES Buffer	
Tris-base	6.06 g
MES	9.76 g
EDTA	0.3 g
(SDS)*	(1 g)*
H_2O	up to 1000 ml

*Not for native gels

4. Remove the tape at the bottom of the gel plate and the comb gently, then insert the gel into the gel running apparatus and add the running buffer.