Revised: October 31, 2023

Product Information

Mini Super^{HT} Pap Pen 2.0
Super^{HT} Pap Pen 2.0

Catalog Number:

Mini Super^{HT} Pap Pen 2.0: 23023
Super^{HT} Pap Pen 2.0: 23024

Unit Size: 1 pen

Size:

Mini Super^{HT} Pap Pen 2.0: 2.5 mm tip, ~400 applications
Super^{HT} Pap Pen 2.0: 4 mm tip, ~800 applications
Number of applications varies depending on barrier size

Storage and Handling

Store at room temperature. The pens contain organic solvents. Avoid skin and eye contact and keep away from open flame. Cap tightly after each use.

Product Description

Super^{HT} Pap Pens are used to create hydrophobic barriers around tissue sections on glass slides to hold staining solutions in place on the slide. This allows the conservation of antibody staining solution or staining of two sections on the same slide with different solutions. The Mini Super^{HT} Pap Pen has a fine point that is useful for separating multiple sections on the same slide. Super^{HT} Pap Pen barriers are insoluble in aqueous buffers, detergents, alcohol and acetone, but can be removed with xylene. Barriers are stable at temperatures up to 120°C.

Directions

Before first use, fill the tip with barrier fluid. Press the tip down on a clean glass microscope slide to open the tip valve (Figure 1). Keep pressing down until the tip becomes saturated with green liquid. Stop pressing as soon as the tip is completely saturated.

Barriers should be created after deparaffinization and rehydration for paraffin sections. For frozen sections, create barriers when slides are dry, before the first buffer incubation step. To create a barrier, dry the area around the section with a cotton swab if necessary. Draw an unbroken circle of Pap Pen fluid around the section (Figure 2). Do not press the valve down while drawing the barrier. Take care to prevent the Pap Pen fluid from touching the tissue section. Let the barrier dry 15-30 seconds before immersing slides or adding buffer. Dry the area immediately outside the barrier if necessary and add just enough buffer to fill the barrier without overflowing. A square of Parafilm® can be placed on top of the section to spread the buffer evenly and prevent evaporation. For overnight incubations, slides should be placed in a humidified chamber to prevent evaporation.

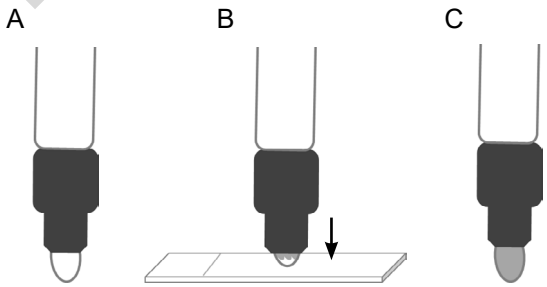


Figure 1. Filling pen tip before first use. A) Pen tip is packaged dry. B) Press tip down against a microscope slide until tip is completely saturated with green liquid (C).

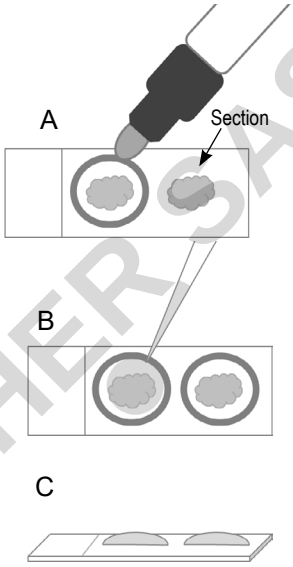


Figure 2. Creating barriers. A) Draw an unbroken circle around sections with Pap Pen. B) Pipette buffer into the barrier. C) Add just enough buffer to fill the barrier without overflowing.

Related Products

Cat. No.	Product
23001	EverBrite™ Mounting Medium
23002	EverBrite™ Mounting Medium with DAPI
23003	EverBrite™ Hardset Mounting Medium
23004	EverBrite™ Hardset Mounting Medium with DAPI
23016	EverBrite™ Hardset Mounting Medium with NucSpot® 640
23005	CoverGrip™ Coverslip Sealant
40083...41038	NucSpot® Nuclear Stains for dead or fixed cells
23007	TrueBlack® Lipofuscin Autofluorescence Quencher
23012	TrueBlack® IF Background Suppressor System (Permeabilizing)
22015	Fixation Buffer
22016	Permeabilization Buffer
22017	Permeabilization and Blocking Buffer (5X)
22010	10% Fish Gelatin Blocking Buffer
22011	Fish Gelatin Powder
22014	30% Bovine Serum Albumin Solution
22002	Tween®-20

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