



NG dART RT-PCR Kit

Cat. No.	size
E0802-01	25 reactions
E0802-02	100 reactions

Storage Conditions: Store at -20°C.

Quality Control:

All preparations are assayed for contaminating endonuclease, exonuclease, nonspecific RNase, single- and double-stranded DNase activities.

References:

- Houts, G.E., Masakau, M., Ellis, C., Beard, D. and Beard, J.W. (1979) *J. Virol.* 29, 517-522.

NG dART RT-PCR Kit is a first strand cDNA synthesis kit convenient for two step RT-PCR. The kit consists of modified reverse transcriptase with improved thermostability (up to 65°C) and processivity. NG dART RT-PCR Kit has easy to use format to save time and limit the possibility of pipetting errors.

NG dART RT-PCR Kit allows to amplify DNA from any RNA with high specificity and sensitivity. **NG dART RT Mix** contains dART reverse transcriptase and RNase Inhibitor preventing from RNases A, B and C. **5 x NG cDNA Buffer** contains optimized for RT reaction buffer and dNTPs.

cDNA synthesis is performed in the first step using either total RNA or poly(A)⁺-RNA primed with oligo(dT), random hexamers primers or reverse gene specific primer. The second step takes place in a separate tube - PCR reaction in which cDNA (as a template) and specific primers are used to amplify double-stranded DNA of interest using high fidelity OptiQ DNA Polymerase.

COMPONENTS OF THE KIT

NG dART RT-PCR Kit	E0802-01	E0802-02
NG dART RT Mix	25 µl	100 µl
5 x NG cDNA Buffer	100 µl	400 µl
Oligo(dT) ₂₀ (50 µM)	25 µl	100 µl
Random hexamers (200 ng/µl)	25 µl	100 µl
dNTPs mix 5 mM each	50 µl	200 µl
OptiQ DNA Polymerase 2.5 U/µl	25 µl	100 µl
10 x Pol Buffer C with MgCl ₂	250 µl	1.0 ml
RNase-free Water	1 ml	4 x 1 ml

This product is developed, designed and sold exclusively for research purposes and in vitro use only.

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First strand cDNA synthesis:

1. Place 5 x NG cDNA Buffer at room temperature, thaw and vortex gently. Visible white precipitate will dissolve and clear buffer is ready for use.
2. Assembly reaction in RNase-free tube as follows:

Component:	Amount:
5 x NG cDNA Buffer	4 μ l
primer*	1 μ l
RNA (10 ng-5 μ g)	x μ l
NG dART RT Mix	1 μ l
RNase-free Water	to 20 μ l

*50 μ M Oligo(dT)₂₀, 200 ng/ μ l random hexamer primer or 10 μ M reverse gene specific primer.

3. Transfer the sample to preheated to appropriate temperature thermal cycler. Incubate as follows:

Oligo(dT)₂₀ primed: 30-60 min at 50°C (or 35-65°C)

Gene specific primed: 30-60 min at 50°C (or 35-65°C)

Random hexamer primed: 25°C for 10 min, followed by 20-50 min at 50°C (or 35-65°C).

NOTE

50°C is suitable temperature for most targets. For G-C rich RNA templates or with complex secondary structure temperature can be increased to 65°C.

4. Terminate the reaction by incubating at 85°C for 5 min.
5. cDNA is ready for PCR, can be used immediately or stored at -20°C. Use 2-5 μ l for 50 μ l PCR.

PCR with OptiTaQ DNA Polymerase:

The final magnesium concentration is 1.5 mM in reaction and in some cases there is a need of titration of magnesium to obtain best results.

1. Mix as follows all reagents in 0.2-0.5 ml tube:

Component:	Amount:
cDNA template	2-5 μ l
10 x Pol Buffer C	5 μ l
dNTPs mix 5 mM each	2 μ l
10 μ M sense primer	1 μ l
10 μ M reverse primer	1 μ l
OptiTaQ DNA Polymerase 2.5 U/ μ l	1 μ l
RNase-free Water	to 50 μ l

50 μ l

2. Mix gently by pipeting.
3. Incubate at 94°C for 3 min, then perform 20-40 cycles of PCR with optimized conditions for your sample (1 min/kb extension time at 68-72°C).
4. Analyze 10-20 μ l of PCR sample by agarose gel electrophoresis.

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