

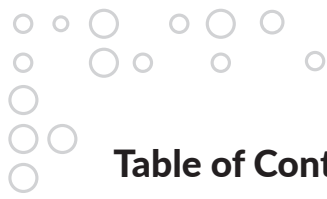
## GeneMATRIX Tissue DNA Purification Kit

Kit for isolation of total DNA from human and animal tissues

● **Cat. no. E3550**

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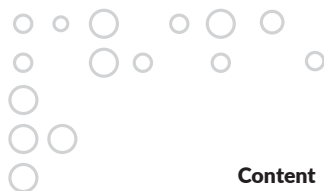
## Introductory Notes

**NOTE 1 • Kit Specification.** The kit is designed for isolation of total DNA (genomic, mitochondrial) from a variety of tissues and biological liquids. To obtain maximum yield we also recommend specialized kits: for isolation of DNA from blood (QuickBlood DNA Purification Kit), cell culture (Cell Culture DNA Purification Kit) or biological traces (Swab Extract DNA Purification Kit and Bio-Trace DNA Purification Kit).

**NOTE 2 • Maximum Sample Amount.** The maximum column binding capacity for DNA is 25 µg. The maximum volume of the column reservoir is 650 µl. One minicolumn enables purification of DNA from up to 25 mg solid tissues or 200 µl liquid tissues.

**NOTE 3 • Kit Compounds Storage.** Once the kit is unpacked, store components at room temperature, with the exception of RNase A and Proteinase K. RNase A should be kept at 2–8°C and Proteinase K at -20°C.

**NOTE 4 • Maintaining Good Working Practice.** All solutions should be kept tightly closed to avoid evaporation and resulting concentration changes of buffer components. To obtain high quality DNA, stick carefully to the protocol provided below. Tissue lysates are very sticky. This can lead to slow lysate filtration through the resin. Therefore it is advisable to check, if lysate and washes passed completely through the resin.



Content	50 preps E3550-01	150 preps E3550-02	Storage/Stability
Buffer T	1.8 ml	5.4 ml	15-25°C
Lyse T	21 ml	63 ml	15-25°C
RNase A (10 mg/ml)	0.12 ml	0.36 ml	2-8°C
Proteinase K (20 mg/ml)	1.2 ml	3.6 ml	-20°C
Sol T	21 ml	63 ml	15-25°C
Wash TX1	30 ml	90 ml	15-25°C
Wash TX2	30 ml	90 ml	15-25°C
Elution	18 ml	54 ml	15-25°C
DNA Binding Columns	50	3 x 50	15-25°C
Protocol	1	1	

### ***Equipment and reagents to be supplied by the experimenter.***

- Microcentrifuge, disposable gloves, sterile pipet tips, sterile 1.5–2 ml tubes, a heating block capable of incubation at 37–70°C.
- [1 M] Dithiothreitol (DTT), ethanol [96–100% v/v], xylene and PBS. To prepare sterile PBS, dissolve 8 g NaCl, 0.2 g KCl, 1.44 g Na<sub>2</sub>HPO<sub>4</sub> and 0.24 g KH<sub>2</sub>PO<sub>4</sub> in 800 ml H<sub>2</sub>O. Adjust pH to 7.4 with HCl. Add H<sub>2</sub>O to 1 liter.

# Protocol

## I. DNA binding spin-columns activation

1. Apply 30  $\mu$ l of activation **Buffer T** onto the **DNA binding spin-column** (do not spin) and keep it at room temperature till transferring lysate to the spin-column (for best results at least 10 min).

- Addition of Buffer T onto the center of the resin enables complete wetting of membranes and maximal binding of DNA.

- The membrane activation should be done before starting isolation procedure.

## II. Sample preparation

### A. Solid tissues

1. Tissue homogenization.
  - a) Grind tissue fragment under liquid nitrogen to a fine powder using previously cooled mortar and pestle. Place sample material (up to 25 mg) in 2 ml Eppendorf tube and centrifuge the powder to the bottom of the tube. Add 350  $\mu$ l of buffer **Lyse T** and suspend the precipitate thoroughly.
    - To obtain high yield of DNA a tissue fragment should be thoroughly grinded to a fine powder.
  - b) Place up to 25 mg of tissue in 2 ml Eppendorf tube. Add 100  $\mu$ l PBS and homogenize the sample using a mechanical homogenizer. Add 250  $\mu$ l of buffer **Lyse T**.
  - c) Cut tissue fragment (up to 25 mg) into small pieces. Place the sample in 2 ml Eppendorf tube and add 350  $\mu$ l of buffer **Lyse T**.
2. Add 2  $\mu$ l of **RNase A** and 20  $\mu$ l of **Proteinase K**. Mix by inverting or vortexing the tube.
3. Incubate at 56°C until the tissue is completely lysed (at least 1–3 h). Mix by inverting or vortexing every 15–30 min.
  - Samples can be lysed overnight, if needed.
4. Follow the point 1. Part III. of the DNA isolation protocol.



## B. Paraffin-embedded tissues

1. Prepare a small section (up to 25 mg) from block of embedded tissue. Place the sample in 2 ml Eppendorf tube.
2. Add 1 ml xylene. Vortex vigorously. Incubate at room temperature for 15 min.
3. Centrifuge for 3 min at 11 000 x g. Remove supernatant by pipetting.
4. Add 1 ml xylene to the pellet, mix by vortexing.
5. Centrifuge for 3 min at 11 000 x g. Remove supernatant by pipetting.
6. Add 1 ml ethanol (96–100%) to the pellet. Mix by vortexing or inverting the tube.
7. Centrifuge for 3 min at 11 000 x g. Remove supernatant by pipetting.
8. Repeat steps 6–7 once.
9. Incubate the open tube at 37°C until the ethanol has evaporated (app. 15 min).
10. Resuspend the tissue pellet in 350 µl of buffer **Lyse T**.
11. Follow the point 2. of A. Solid tissues protocol.

## C. Formalin-fixed tissues

1. Wash tissue sample twice with PBS to remove fixative. Discard PBS.
2. Cut tissue fragment (up to 25 mg) into small pieces. Place the sample in 2 ml Eppendorf tube and add 350 µl of buffer **Lyse T**.
3. Follow the point 2. of A. Solid tissues protocol.

## D. Liquid tissues/body fluids

(blood, saliva, plasma, serum, brain-spinal cord liquid among others).

1. Add 2 µl **RNase A** to 200 µl liquid sample. Mix thoroughly by vortexing the tube.
  - For sample volumes less than 200 µl, add PBS to adjust the volume to 200 µl.
2. Incubate for 5 min at room temperature.
3. Add 10 µl **Proteinase K**.
4. Follow the point 1. Part III. of the DNA isolation protocol.

## E. Cultured cells

1. Centrifuge the cell culture (up to  $5 \times 10^6$  cells) in the 1.5–2 ml Eppendorf tube for 2 min at 1000 x g.
2. Carefully discard the supernatant. Add to the pellet 200  $\mu$ l of **Lyse T** buffer and 2  $\mu$ l of **RNase A**. Suspend the cells thoroughly by vortexing for 20 sec.
3. Incubate for 5 min at room temperature.
4. Add 10  $\mu$ l **Proteinase K**.
5. Follow the point 1. Part III. of the DNA isolation protocol.

## F. Rodent tails

1. Cut up to 1.2 cm-piece of mouse tail or up to 0.6 cm-piece of rat tail into the 2 ml Eppendorf tube. Add 350  $\mu$ l of buffer **Lyse T**.
2. Add 2  $\mu$ l of **RNase A** and 20  $\mu$ l of **Proteinase K**. Mix by vortexing.
3. Incubate at 56°C until the tissue is completely lysed. Mix by vortexing every 1 h or use a shaking water bath.
  - *Samples can be lysed overnight.*
4. Vortex for 15 sec. Centrifuge for 3 min at 11 000 x g. Transfer the supernatant into a new tube.
  - *This step removes residual bones and hair.*
5. Add 350  $\mu$ l of buffer **Sol T**. Add 350  $\mu$ l of ethanol (96–100%). Mix thoroughly by vortexing.
6. Follow the point 6. Part III. of the DNA isolation protocol.

## G. Hair

1. Cut off the hair roots from the hair sample (up to 100 roots or 25 mg). Place them in the 2 ml Eppendorf tube. Add 350  $\mu$ l of buffer **Lyse T**, 20  $\mu$ l of 1M DTT and 20  $\mu$ l of **Proteinase K**. Mix by vortexing.
  - *If the hair sample doesn't contain the roots cut the hair stems into short pieces not longer than 0.5 cm.*
  - *The hair stem is the dead part of hair that contain small quantities of degraded DNA. The recommended amplicon length for PCR analysis of DNA from the hair stems is <200 bp.*
2. Incubate at 56°C until the hair sample is completely lysed (6–8 h or overnight).
3. Mix by vortexing every 1–2 h or use a shaking water bath.
4. Follow the point 1. Part III. of the DNA isolation protocol.



## H. Insects

### 1. Insect homogenization.

a) Grind insects under liquid nitrogen to a fine powder using previously cooled mortar and pestle. Place the powder (up to 50 mg) in 2 ml Eppendorf tube. Centrifuge the sample to the bottom of the tube. Add 350  $\mu$ l of buffer **Lyse T** and suspend the precipitate thoroughly.

○ *To obtain high yield of DNA a tissue fragment should be thoroughly grinded to a fine powder.*

b) Place up to 50 mg insects in 2 ml Eppendorf tube. Add 100  $\mu$ l PBS and homogenize the sample using a mechanical homogenizer. Add 250  $\mu$ l of buffer **Lyse T**.

### 2. Follow the point 2. of A. Solid tissues protocol.

## I. Urine.

1. Add 2 ml of urine to the 2 ml Eppendorf tube.

2. Centrifuge urine in microcentrifuge for 2 min at 6 000 x g.

3. Carefully discard the supernatant without disturbing the pellet. Add to the pellet 350  $\mu$ l **Lyse T** buffer and 10  $\mu$ l **Proteinase K**.

4. Vortex for 15 sec.

5. Incubate for 60 min at 56°C, mix by inverting the tube every 15 min.

6. Follow the point 1. Part III. of the DNA isolation protocol.

## III. DNA isolation

1. Add 200  $\mu$ l of buffer **So! T** (D. Liquid tissues, E. Cultured cells) or 350  $\mu$ l of buffer **So! T** (A. Solid tissues, B. Paraffin-embedded tissues, C. Formalin-fixed tissues, G. Hair, H. Insects, I. Urine) and mix thoroughly by vortexing or several-fold inverting.


2. Incubate for 10 min at 70°C.

3. Add 200  $\mu$ l of ethanol (96–100%) (D. Liquid tissues, E. Cultured cells) or 350  $\mu$ l of ethanol (96–100%) (A. Solid tissues, B. Paraffin-embedded tissues, C. Formalin-fixed tissues, G. Hair, H. Insects, I. Urine).

4. Mix thoroughly by several times inverting the tube.

5. Centrifuge for 1 min at 12 000 x g.



- 
6. Transfer the whole lysate (D. Liquid tissues, E. Cultured cells) or 600  $\mu$ l of supernatant (A. Solid tissues, B. Paraffin-embedded tissues, C. Formalin-fixed tissues, F. Rodent tails, G. Hair, H. Insects, I. Urine) to the **DNA binding spin-column**, placed in the collection tube.
  7. Centrifuge **DNA binding spin-column** for 1 min at 11 000 x g.
    - Continue centrifugation at maximum speed, if not all of the lysate passed through the column.
  8. Remove the spin-column, discard flow-through and place back spin-column in the collection tube. In cases D. Liquid tissues and E. Cultured cells proceed with step 11.
  9. Transfer the remaining supernatant (A. Solid tissues, B. Paraffin-embedded tissues, C. Formalin-fixed tissues, F. Rodent tails, G. Hair, H. Insects, I. Urine) to the **DNA binding spin-column**, placed in the collection tube. Repeat centrifugation for 2 min at 11 000 x g to pass completely the lysate through the resin.
    - Continue centrifugation, if not all of the lysate passed through the column.
  10. Take out **DNA binding spin-column**, discard flow-through and place back spin-column in the collection tube.
  11. Add 500  $\mu$ l of **Wash TX1** buffer and centrifuge for 1 min at 11 000 x g.
  12. Take out **DNA binding spin-column**, discard flow-through and place back spin-column in the collection tube.
  13. Add 500  $\mu$ l of **Wash TX2** buffer and centrifuge for 1 min at 11 000 x g.
  14. Spin down at 11 000 x g for 1 min to remove traces of **Wash TX2** buffer.
  15. Place **DNA binding spin-column** in a new collection tube (1.5–2 ml) and add 50–150  $\mu$ l of **Elution** buffer to elute bound DNA.
    - Addition of the elution buffer directly onto the center of the resin improves DNA yield. To avoid transferring traces of DNA between the spin-columns do not touch the spin-column walls with the micro-pipette.
    - In order to improve the efficiency of the elution genomic DNA from membrane, Elution buffer can be heated to a temperature of 80°C.
  16. Incubate **DNA binding spin-column**/collection tube assembly for 2 min at room temperature.
  17. Centrifuge for 1 min at 11 000 x g.
  18. Remove spin-column, cap the collection tube. DNA is ready for analysis/manipulation. It can be stored either at 2–8°C or at -20°C.



## Appendix 1:

### Detection of *Mycobacterium tuberculosis* in sputum or bronchoalveolar lavage

1. Add 1 volume of NALC-NaOH solution (2% NaOH, 1.45% sodium citrate, 0.5% N-acetyl-L-cysteine) to 200–500  $\mu$ l of sputum or bronchoalveolar lavage.
  - *To prepare NALC-NaOH solution dissolve: 2 g NaOH, 1.45 g sodium citrate, 0.5 g N-acetyl-L-cysteine. Add sterile distilled water to 100 ml.*
2. Mix by vortexing and incubate for 20 min at room temperature. Mix by vortexing or inverting every 5 min.
3. Adjust the volume to 25 ml with sterile distilled water.
4. Centrifuge for 30 min at 4 000 x g. Discard the supernatant.
5. Resuspend the pellet in 0.5–1 ml of buffer **Lyse T**.
6. Transfer 200  $\mu$ l of the sample to a new microcentrifuge tube.
7. Add 20  $\mu$ l **Proteinase K**. Mix by vortexing or inverting the tube.
8. Incubate for 1 hr at 56°C. Mix by vortexing or inverting every 15 min.
9. Follow the point 1. Part III. DNA isolation according to protocol for: D. Liquid tissues, E. Cultured cells.

# Safety Information

## Buffer T

### Danger



**H314** Causes severe skin burns and eye damage.

**P280** Wear protective gloves/protective clothing/eye protection/face protection.

**P301+P330+P331** If swallowed: Rinse mouth. Do not induce vomiting.

**P303+P361+P353** If on skin (or hair): take off immediately all contaminated clothing. Rinse skin with water [or shower].

**P305+P351+P338** If in eyes: rinse cautiously with water for several minutes. Remove contact lenses, if present and easy to do. Continue rinsing.

**P310** Immediately call a poison center/doctor.

**P405** Store locked up.

## Lyse T

### Warning



**H319** Causes serious eye irritation.

**P280** Wear protective gloves/protective clothing/eye protection/face protection.

**P305+P351+P338** If in eyes: rinse cautiously with water for several minutes. Remove contact lenses, if present and easy to do. Continue rinsing.

**P337+P313** If eye irritation persists: Get medical advice/ attention.

## Proteinase K

### Danger



**H334** May cause allergy or asthma symptoms or breathing difficulties if inhaled.

**P261** Avoid breathing vapours/spray.

**P304+P340** If inhaled: remove person to fresh air and keep comfortable for breathing.

**P342+P311** If experiencing respiratory symptoms: call a poison center or doctor/ physician.

## Sol T

### Warning



**H302+H332** Harmful if swallowed or if inhaled.

**H315** Causes skin irritation.

**H319** Causes serious eye irritation.

**P261** Avoid breathing vapours/spray.

**P280** Wear protective gloves/protective clothing/eye protection/face protection.

**P301+P312** If swallowed: call a poison center/ doctor if you feel unwell.

**P304+P340** If inhaled: remove person to fresh air and keep comfortable for breathing.

**P305+P351+P338** If in eyes: rinse cautiously with water for several minutes. Remove contact lenses, if present and easy to do. Continue rinsing.

**P333+P313** If skin irritation or rash occurs: get medical advice/attention.

**P337+P313** If eye irritation persists: get medical advice/ attention.

**EUH208** Contains ethylenediammonium dichloride. May produce an allergic reaction.

## Wash TX1

### Warning



**H226** Flammable liquid and vapour.

**H302+H332** Harmful if swallowed or if inhaled.

**H315** Causes skin irritation.

**H319** Causes serious eye irritation.

**P210** Keep away from heat, hot surfaces, sparks, open flames and other ignition sources. No smoking.

**P280** Wear protective gloves/protective clothing/eye protection/face protection.

**P301+P312** If swallowed: call a poison center/ doctor if you feel unwell.

**P302+P352** If on skin: wash with plenty of water.

**P304+P340** If inhaled: remove person to fresh air and keep comfortable for breathing.

**P305+P351+P338** If in eyes: rinse cautiously with water for several minutes. Remove contact lenses, if present and easy to do. Continue rinsing.

## Wash TX2

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### Danger



**H225** Highly flammable liquid and vapour.

**H319** Causes serious eye irritation.

**P210** Keep away from heat, hot surfaces, sparks, open flames and other ignition sources. No smoking.



**P280** Wear protective gloves/protective clothing/eye protection/face protection.

**P305+P351+P338** If in eyes: rinse cautiously with water for several minutes. Remove contact lenses, if present and easy to do. Continue rinsing.

**P403+P235** Store in a well-ventilated place. Keep cool.

**P337+P313** If eye irritation persists: get medical advice/ attention.

**SELECTION OF THE KITS  
DEPENDING ON THE TYPE  
OF ISOLATED MATERIAL**

		ISOLATION OF DNA																									
		E3600	E3685	E3640	E3680	E3610	E3645	E3660	E3655	E3625	E3620	E3695	E3635	E3600	E3665	E3615	E3670	E3675	E3630	E3650	E3651						
		MI-CELLULLA DNA <sup>2</sup>	GRAM PLUS & YEAST GENOMIC DNA	AGAROSE - OUT DNA	BACTERIAL & YEAST GENOMIC DNA	BIO - TRACE DNA	BASIC DNA	BONE DNA	CELL CULTURE DNA	FOOD EXTRACT DNA	PCR / DNA CLEAN-UP	PLANT & FUNGI DNA	AGROBACTERIUM PLASMID DNA	PLASMID MINIPREP DNA	QUICK BLOOD DNA	SHORT DNA CLEAN-UP	SOIL DNA	STOOL DNA	SWAB-EXTRACT DNA	TISSUE DNA	TISSUE & BACTERIAL DNA						
		AVAILABLE NUMBER OF ISOLATION (PREPS)																									
		50 150	25 100	50 150	50 150	25 100	50 150	25 05	50 150	25 100	50 150	50 150	50 150	50 150	50 150	25 100	50 100	50 100	25 100	50 150	50 150						
DNA	GENOMIC	BACTERIA	●		●																	●					
		YEAST	●		●																						
		CELL CULTURE								●													●	●			
		PLANT																									
		FUNGI																									
		PLANT RICH IN POLYSACCHARIDES <sup>1</sup>																									
		BLOOD																									
		SOIL																									
		STOOL																									
		SWAB																									
		ANIMAL TISSUES																									
		FFPE TISSUE SECTIONS																									
		RODENT TAILS																									
		HAIR																									
		INSECTS																									
		URINE																									
		BONE																									
	BIOLOGICAL TRACES																										
	FOOD																										
	PLASMID	BACTERIA																									
YEAST																											
ISOLATION FROM AGAROSE GELS																											
PURIFICATION OF PCR PRODUCTS / DNA AFTER ENZYMATIC REACTIONS		●																									

All kits contain buffers WASH in ready to use form

1. Additionally required lyse CT buffer (E0324)

2. Kit for creation of emulsions and subsequent DNA purification.

**SELECTION OF THE KITS  
DEPENDING ON THE TYPE  
OF ISOLATED MATERIAL**

		ISOLATION OF RNA							
		E3700	E3934	E3936	E3938	E3939	E3933		
		RNA EXTRACOL 2	UNIVERSAL BLOOD RNA	HUMAN BLOOD RNA	UNIVERSAL RNA	UNIVERSAL RNA/miRNA	FFPE RNA Purification Kit		
		PREPS							
		25 100	25	25	25 100	25 100	25 100		
RNA	TOTAL RNA LONGER THAN 200 BASES	ANIMAL TISSUE				●	●		
		PLANT TISSUE				●	●		
		BACTERIA				●			
		YEAST				●			
		CELL CULTURE				●	●		
		HUMAN BLOOD	FRESH	●	●	●	●		
			FROZEN <sup>1</sup>		●				
		ANIMAL BLOOD	FRESH	●	●				
	FROZEN <sup>1</sup>			●					
	miRNA OR TOTAL RNA	ANIMAL TISSUE	●				●		
		FFPE TISSUE SECTIONS						●	
		PLANT TISSUE	●				●		
		CELL CULTURE	●				●		
		BACTERIA	●						
		YEAST	●						
BLOOD/LEUKOCYTES		●							
PURIFICATION OF RNA AFTER ENZYMATIC REACTIONS					●	●			
ON-COLUMN DNase DIGESTION			●		●				

All kits contain buffers WASH in ready to use form

1. Frozen with the addition of Lyse Blood buffer (included in kit).
2. Phenol-based reagent for isolation RNA.

○ **GeneMATRIX is synthetic, new generation DNA- and RNA-binding membrane, selectively binding nucleic acids to composite silica structures.**

Novel binding and washing buffers are developed to take full advantage of GeneMATRIX capacity, yielding biologically active, high-quality nucleic acids. Matrix is conveniently pre-packed in ready-to-use spin-format. Unique chemical composition of the matrixes along with optimized construction of spin-columns improve the quality of final DNA or RNA preparation. To speed up and simplify isolation procedure, the key buffers are colour coded, which allows monitoring of complete solution mixing and makes purification procedure more reproducible.

As a result, we offer kits, containing matrixes and buffers that guarantee rapid, convenient, safe and efficient isolation of ultrapure nucleic acids. Such DNA or RNA can be directly used in subsequent molecular biology applications, such as: restriction digestion, dephosphorylation, kinasing, ligation, protein-DNA interaction studies, sequencing, blotting, in vitro translation, cDNA synthesis, hybridization among others. Additional advantage is reproducibility of matrix performance, as component preparation is carried at Eurx Ltd.

○ **GeneMATRIX Tissue DNA Purification Kit is designed for rapid purification of total DNA (genomic, mitochondrial) from a variety of tissues and biological liquids. Purified DNA is free of contaminants, such as: RNA, proteins, lipids, dyes, detergents, organic inhibitors of enzymatic reactions, buffers, salts, divalent cations, among others.**

Sample is proteolytically lysed in the presence of special buffer, aiding tissue and cells desintegration. Further, Proteinase K digests cellular proteins, including stripping-off DNA of all bound proteins, among them nucleases. Optimized buffer and ethanol are added to provide selective conditions for DNA binding during brief centrifugation, while contaminants pass through the GeneMATRIX resin in the spin-column. Traces of contaminants remaining

on the resin are efficiently removed in two wash steps. High-quality cellular DNA is then eluted in low salt buffer, e.g.: Tris-HCl, TE or water. Isolated DNA is ready for downstream applications without the need for ethanol precipitation



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