

For use in automated high throughput applications including sample preparation and radiometric filter-binding assays

For research use only Single use only



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Introduction

MultiScreen_{HTS} 384-well plates are non-sterile, single use filter plates that can be used to perform high throughput biological assays or to prepare samples for subsequent analysis. The filter plate can be used in a variety of biological assays including receptor-ligand binding (RLB) assays, assays that measure enzyme activity (e.g., kinases, phosphatases, proteases, etc.), and general sample prep.

The MultiScreen_{HTS} 384-well plate is designed to be compatible with robotic workstations for automated manipulation and can be used with either vacuum or centrifugal filtration modes. These filter plates are compatible with 384-well receiver plates for the collection and analysis of filtered samples.





Materials Required

The user must supply the following materials in order to use the MultiScreen_{HTS} 384-well filter plate:

Reagents:

Application-appropriate aqueous buffers, biological components, and other assay materials

Equipment:

- Manual pipettors or robotic liquid handling systems for transferring of liquid into filter plates
- Vacuum manifold (Millipore cat. no. MSVM HTS 00) or centrifuge with a swinging bucket rotor and multiwell plate carrier
- Solid bottom 384-well assay plates (if collecting filtrate)

Chemical Compatibility

R= Resistant, N= Non-Resistant, T= User should test in application

	Com	Compatibility	
Chemical	Membrane	Glass/Paper	
Acids			
Acetic Acid (Glacial)	Ν	Ν	
Acetic Acid (25%)	R	R	
Sulfuric Acid (90%)	N	Ν	
Scintillants			
MicroScint-O TM Scintillant	R	R	
MicroScint [®] 40 Scintillant	R	R	
Opti-Fluor [®] Scintillant	R	R	
OptiPhase [™] Supermix [™] Scintilla	ant N	R	
Ultima Gold [™] Scintillant	R	R	
Organic Solvents			
Acetone	Ν	Ν	
Acetonitrile	Ν	Ν	
Tetrahydrofuran	Ν	Ν	
Diethyl Ether	Ν	Ν	
DMSO	Ν	Ν	
Glycerol	R	R	
Methanol	Т	Т	
Aqueous Solvents			
Sodium Dodecyl Sulfate (SDS)	R	Т	
Tween [®] Surfactant	R	Т	
Phosphate Buffered Saline (PBS)) R	R	
1% Phosphoric Acid	R	R	
1 N HCl	R	R	
1 N NaOH	R	R	
Acetonitrile (35%)	Ν	Ν	
DMSO (5%)	R	R	

Usage Guidelines

- The MultiScreen_{HTS} 384-well filter plate can be operated either in a vacuum or centrifugal pressure mode at temperatures between 4 °C and 37 °C. Recommended vacuum pressure is 8–15" Hg for glass fiber plates and membrane plates or 4–8" Hg for PH plates. Recommended centrifugal force is 500–1000 × g.
- In either vacuum or centrifugal mode, samples can be collected in a solid bottom receiver (assay) plate. In vacuum mode, sample or (wash) buffer(s) can be collected into a solid bottom receiver plate or transferred directly to waste.
- Although the MultiScreen_{HTS} 384-well plate has been validated for use with extended incubations of aqueous media and scintillants, care should be taken to protect working surfaces from contamination in case of leakage.
- When doing radiometric in-plate analysis with the Wallac MicroBeta[®] TriLux liquid scintillation counter, plate carriers should be taped.
- Extended incubations with solutions containing high levels of surfactant may lead to drip out; test the plate before use in experiments.
- The MultiScreen_{HTS} 384-well filter plate with glass fiber is compatible with polyethyleneimine (0.5% PEI) pretreatment. **Do not allow PEI to dry!** For those applications requiring this step, wash twice with binding buffer to complete preparation of plates after initial pretreatment for 2–16 hours at 4 °C.
- Dry glass fiber plates after filtration if adding scintillant. Do not dry PH or DEAE plates if adding scintillant. After adding scintillant, wait at least 1 hour before reading to allow scintillant to fully coat the filter.
- Cover the plate with the lid while incubating or use some other means to reduce evaporation, especially if incubating for more than 1 hour or at elevated temperatures.
 - If incubating a reaction in PH plates, do not exceed 80 µL of sample per well.

Limitations

Operating Temperature Range	4–37 °C
Maximum Drying Temperature	50 °C
Maximum Vacuum Pressure	20" Hg
Maximum Centrifugal Force	$1000 \times g$

Procedure

The following sections outline the use of the MultiScreen_{HTS} 384-well filter plate.

Sample Addition

Pipette the appropriate amount of test sample, 1 to 100 μ L, into each well of the filtration plate. For best results when adding multiple solutions to the well, add the solution with the largest volume first, and end by adding the solution containing the smallest volume. Well volume limit is 80 μ L for PH plates.

In-Plate Incubation

The MultiScreen_{HTS} 384-well filter plate is designed to permit in-plate incubations of various liquids, thereby eliminating the need to perform assays in a separate plate prior to transferring the sample to the filter plate. Due to the low volumes typically incubated in the MultiScreen_{HTS} 384-well filter plate (20–100 μ L), it is recommended that the plate lid be used during extended incubations or incubations that are carried out at elevated temperatures (e.g., at or above 37 °C). Well volume limit is 80 μ L for PH plates.

Sample Filtration

Samples can be filtered using either centrifugal or vacuum modes. In centrifugal mode, a 384-well receiver plate is positioned under the filter plate to collect the filtrate. Vacuum filtration should be performed using the MultiScreen_{HTS} Vacuum Manifold (Millipore cat. no. MSVM HTS 00). Please see the MultiScreen_{HTS} Vacuum Manifold User Guide (P36530) for specific guidelines. In vacuum mode, the filtrate is either collected in a receiver plate or it is transferred directly to waste.

Filtrate Collection

The filter plate is designed to be used in conjunction with an SBS/ANSI solid bottom 384-well receiver (assay) plate. When stacked on top of a 384-well receiver plate, the Multi-Screen_{HTS} 384-well plate will direct the contents of each well into the corresponding well in the receiver plate.

For centrifugation, stack the filter plate on top of a receiver plate and then place the assembly into the plate carrier of any standard centrifuge with swinging bucket rotors.

In vacuum mode, stack the filter plate on top of the receiver plate and place the assembly in the base of the vacuum manifold. The collar fits over the assembly. See the MultiScreen_{HTS} Vacuum Manifold User Guide for details.

NOTE: Optimal vacuum levels can vary among assays. Typically, the lowest vacuum setting that gives the desired performance should be used.

Washing of Samples to Waste

For optimal performance during multiple washes of retentate sample (i.e., flow to waste), vacuum pressures of 12–15" Hg are recommended, depending on filter type.

Automated Operation Guidelines

The MultiScreen_{HTS} 384-well filter plate is designed to be used with automated liquid handling systems and robotic workstations that are compatible with ANSI/SBS standard 384-well plates. Integration of vacuum filtering capability may require some customization or be impossible on some systems. For the latest automation protocols and robotic workstation-specific programs and instruction, please see: http://www.millipore.com/automation.

Radiometric Assay Counting Guidelines

Follow the guidelines below for in-plate measurement of radioactivity. For information on receptor ligand binding assays or kinase assays, go to www.millipore.com/publications.

Whole Plate Scintillation Counting

Radioactivity (scintillation counting) can be measured directly in the MultiScreen_{HTS} 384-well filter plate. The design allows the plate to be analyzed from the top (top counting), from the bottom (bottom counting), or from both sides simultaneously (coincidence counting). For best signal to noise ratios using the Wallac MicroBeta TriLux liquid scintillation counter, count in the coincidence mode. Scintillation counting should be performed after at least one hour of exposure to the liquid scintillant cocktail. In some instances, longer incubations (up to 24 hours) may result in even higher counting efficiencies. Because different assays have different counting criteria and levels of signal, optimum counting mode and time should be determined by the operator.

Radioisotope Guidelines

Low energy isotopes such as ¹⁴C, ³H, ¹²⁵I, ³³P, and ³⁵S require the use of 5–10 μ L of scintillation cocktail per well. For optimal results and consistency, the scintillation cocktail should be allowed to mix with the sample for at least an hour before counting. This time is required to allow for complete mixing of the radiolabeled materials into the scintillation cocktail.

Specifications

Materials of construction:

Base plate	Styrene Acrylonitrile (SAN)
	top: white; bottom: clear
Plate lid	Polystyrene
Support	Polyester (glass fiber and filter paper only)
Membrane	PVDF

See Product Ordering Information table for filter types.

Dimensions:

Plate assembly length	127.8 mm
Plate assembly width	85.5 mm
Plate assembly depth (without lid)	14.4 mm
Plate assembly depth (with lid)	16.9 mm
Approximate filter surface area	0.09 cm^2
Well depth	approx. 10.9 mm
Recommended sample volume per well:	20–100 µL
Filtration parameters:	
Recommended vacuum pressure	8–15" Hg for glass fiber and membrane plates
	4–8" Hg for PH plates
Recommended centrifugal force	$500-1000 \times g$

Product Ordering Information

Catalog Numbers	Membrane/Filter Type	Qty
MZHV N0W 10	Hydrophilic 0.45 μm Durapore [®] PVDF	10/pk
MZHV N0W 50	Hydrophilic 0.45 μm Durapore PVDF	50/pk
MZFB N0W 10	Glass Fiber B	10/pk
MZFB N0W50	Glass Fiber B	50/pk
MZFC N0W 10	Glass Fiber C	10/pk
MZFC N0W 50	Glass Fiber C	50/pk
MZPH NOW 10	Phosphocellulose	10/pk
MZPH NOW 50	Phosphocellulose	50/pk

Accessories

Description	Catalogue No.
MultiScreen _{HTS} Vacuum Manifold	MSVM HTS 00
Chemical Duty Pump	
115 Volts, 60 Hz 220 Volts, 50 Hz	WP61 115 60 WP61 220 50
Spout Liner for Scintillation Counting	MZMB NCL 50
Plate Sealing Tape, 100 sheets	
Clear	MATA HCL 00
White	MATA H0P 00
Vacuum Flask, 1L	XX10 047 05

Technical Assistance

For more information, contact the Millipore office nearest you. In the U.S., call **1-800-MILLIPORE** (1-800-645-5476). Outside the U.S., see your Millipore catalogue for the phone number of the office nearest you or go to our web site at www.millipore.com/offices for up-to-date worldwide contact information. You can also visit the tech service page on our web site at http://www.millipore.com/techservice.

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